

BLEACH&STAIN 15 marker multiplexed imaging in 3098 human carcinomas revealed six major PD-L1 driven immune phenotypes with distinct spatial orchestration.

Introduction and Objectives

Highly multiplexed fluorescence (IHC) enables immunohistochemistry quantification of checkpoints immune such as PD-L1 (programmed cell death ligand 1), PD-1 (programmed cell death protein 1) or CTLA-4 (cytotoxic Tlymphocyte-associated protein 4) in the tumor microenvironment (TME). However, framework to assess the spatial orchestration of these markers and immune cells has yet to be established.

Materials & Methods

To study the impact of PD-L1, PD-1 and CTLA-4 expression on the TME and patient's outcome, a framework for checkpoint automated immune quantification on tumor and immune cells was established and validated. Automated immune checkpoint quantification was facilitated by incorporating three different deep learning steps for the analysis of 44 different human carcinomas from 3098 tumor specimens using a bleach & stain 15-marker multiplex fluorescence IHC panel was used for this study (i.e., PD-L1, PD-1, CTLA-4, panCK, CD68, CD163, CD11c, iNOS, CD3, CD8, CD4, FOXP3, CD20, Ki-67, CD31).



A high intratumoral macrophage density, a high degree of T-cell exclusion, a high PD-1 expression on T-cells, and a low CTLA-4 expression level on T-cells was significantly associated with the inflamed PD-L1 immune phenotype (p<0.001 each). A high M1 macrophage (CD68+iNOS+CD163-) density was linked to the non-inflamed PD-L1⁺ macrophage phenotype (p=0.048). All other macrophage and DC subsets (identified by CD68, CD163, CD11c, iNOS) as well as T-cell subsets (identified by CD3, CD8, CD4, FOXP3) were linked to the inflamed PD-L1 immune phenotypes (p<0.001 each).

Cycle 1	Cycle 2	Cycle 3	Cycle 4	Cycle 5	
CD68	CTLA-4	panCK	FOXP3	Ki-67	
PD-L1	CD11c	CD3	CD8	CD31	
PD-1	CD163	INOS	CD4	CD20	

15+1 BLEACH&STAIN

validated We developed and a novel immunohistochemistry approach that enables high throughput 15 marker staining. Bleeding, crosstalk, or leftover staining was not detected across the 5 staining cycles.



Elena Bady¹, Katharina Möller¹, Tim Mandelkow¹, Ronald Simon¹, Maximilian Lennartz¹, Claudia Hube-Magg¹, Guido Sauter¹, Niclas C. Blessin¹ ¹Institute of Pathology, University Medical Center Hamburg-Eppendorf, Germany



Conflicts of interest: The PD-L1, CTLA-4, panCK, CD4, Ki-67, CD31, and CD20 antibody clones were provided by MS Validated Antibodies GmbH (owned by a family member of GS)

In the inflamed PD-L1⁺ patients (G1.1), spatial analysis revealed that an elevated intratumoral CD68+CD163+ M2 macrophages as well as CD11c+ dendritic cell infiltration (p<0.001 each) was associated with a high (CD3⁺CD4[±]CD8[±]FOXP3[±]) T-cell exclusion and a high PD-1 expression on T-cells (p<0.001 each).

In breast cancer, PD-L the intensity on cells tumor showed a higher predictive performance for overall survival with an area under receiver operating curves (AUC) of 0.72 (p<0.0001) than percentage the of PD-L1⁺ tumor cells (AUC: 0.54)

- inflamed TME and high spatial T-cell exclusion.
- \geq 6 major PD-L1 phenotypes were identified ranging from:
 - an inflamed PD-L1⁺ tumor cell phenotype with a **spatial T-cell exclusion** (G1.1)
 - to a non-inflamed PD-L1⁺ immune cell phenotype showing a particular poor prognosis (G2.2)
- to a non-inflamed PD-L1⁻ negative phenotype (G3.2).



Spatial orchestration of immune cells across the PD-L1 im mune phenotypes



Conclusions

> The highly multiplexed fluorescence IHC BLEACH&STAIN framework for automated PD-L1, PD-1 and CTLA-4 assessment revealed that PD-L1 positive tumor samples can be also associated with a non-