

BLEACH&STAIN 15 marker multiplex fluorescence immunohistochemistry revealed six major PD-L1 immune phenotypes with distinct spatial orchestration.

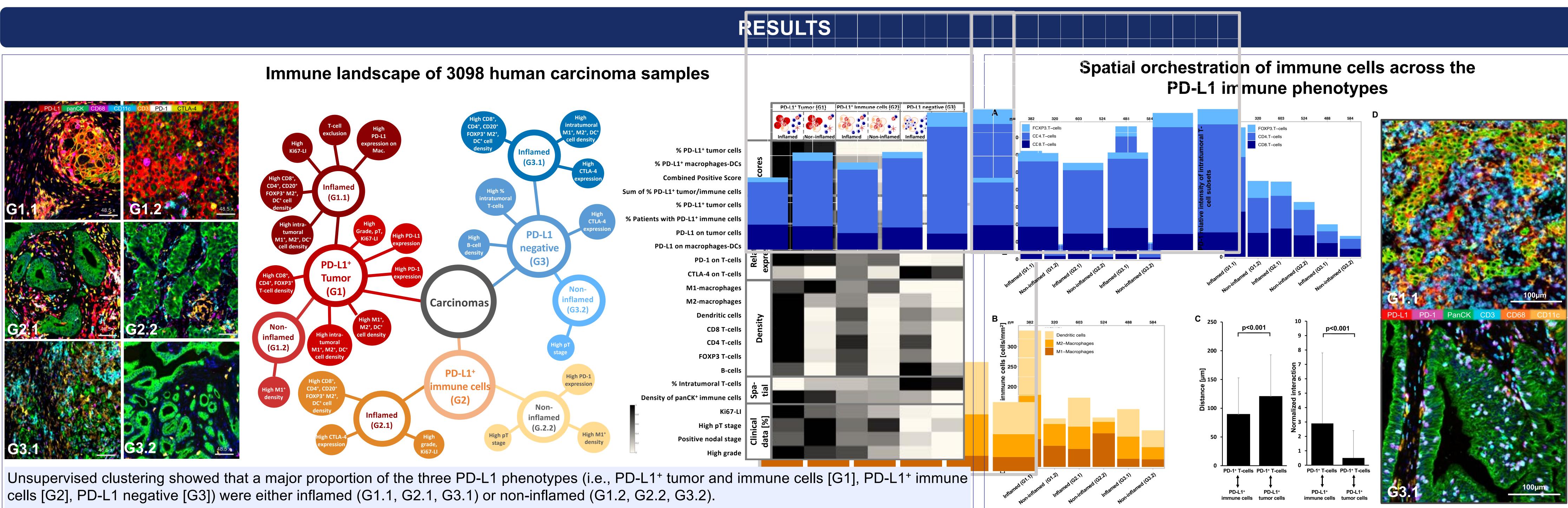
Niclas C. Blessin¹, Elena Bady¹, Tim Mandelkow¹, Cheng Yang¹, Jonas B. Raedler^{1,2}, Ronald Simon¹, Eike Burandt¹, Doris Höflmayer¹, Guido Sauter¹, Katharina Möller¹, Sören A. Weidemann¹ ¹Institute of Pathology, University Medical Center Hamburg-Eppendorf, Germany, ²College of Arts and Science, Boston University, Massachusetts, USA

Introduction and Objectives

fluorescence multiplexed immunohistochemistry enables (IHC) quantification of immune checkpoints such as PD-L1 (programmed cell death ligand 1), PD-1 (programmed cell death protein 1) or CTLA-4 (cytotoxic T-lymphocyteassociated protein 4) in the tumor However, (TME). microenvironment spatial framework the of these markers and orchestration immune cells has yet to be established.

Materials & Methods

To study the impact of PD-L1, PD-1 and CTLA-4 expression on the TME and patient's outcome, a framework for checkpoint automated immune quantification on tumor and immune cells was established and validated. Automated immune checkpoint quantification was facilitated by incorporating three different deep learning steps for the analysis of 44 different human carcinomas from 3098 tumor specimens using a bleach & stain 15-marker multiplex fluorescence IHC panel was used for this study (i.e., PD-L1 PD-1, CTLA-4, panCK, CD68, CD163, CD11c, iNOS, CD3, CD8, CD4, FOXP3, CD20, Ki67, CD31).



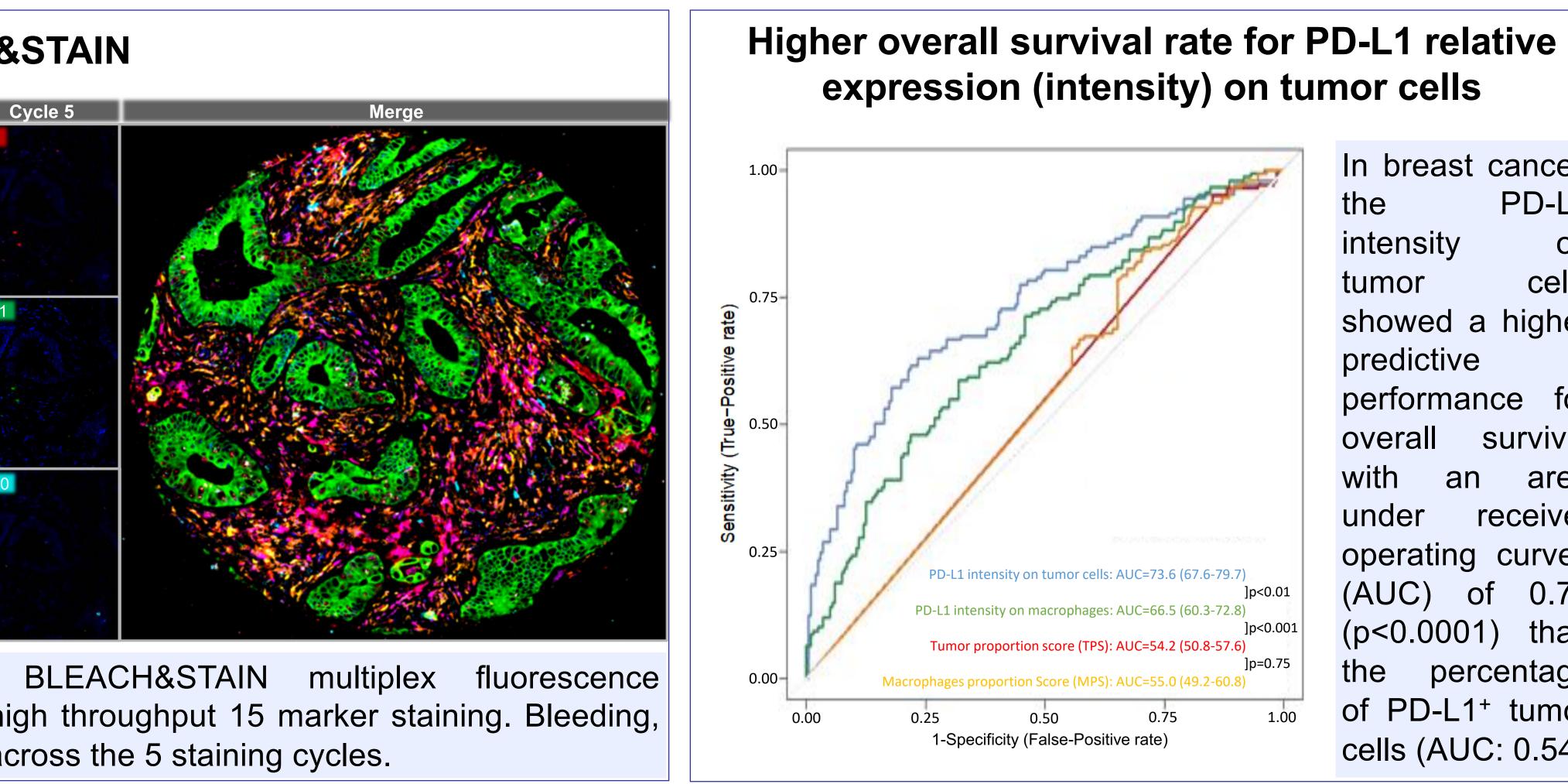
A high intratumoral macrophage density, a high degree of T-cell exclusion, a high PD-1 expression on T-cells, and a low CTLA-4 expression level on T-cells was significantly associated with the inflamed PD-L1 immune phenotype (p<0.001 each). A high M1 macrophage (CD68⁺iNOS⁺CD163⁻) density was linked to the non-inflamed PD-L1⁺ macrophage phenotype (p=0.048). All other macrophage and DC subsets (identified by CD68, CD163, CD11c, iNOS) as well as T-cell subsets (identified by CD3, CD8, CD4, FOXP3) were linked to the inflamed PD-L1 immune phenotypes (p<0.001 each).

Cycle 1	Cycle 2	Cycle 3	Cycle 4	Cycle 5	
CD68	CTLA-4	panCK	FOXP3	Ki67	
PD-L1	CD11c	CD3	CD8	CD31	
PD-1	CD163	inos	CD4	CD20	

15+1 BLEACH&STAIN

We developed and validated a novel immunohistochemistry approach that enables high throughput 15 marker staining. Bleeding, crosstalk, or leftover staining was not detected across the 5 staining cycles.





Conflicts of interest: The PD-L1, CTLA-4, panCK, CD4, Ki67, CD31, and CD20 antibody clones were provided by MS Validated Antibodies GmbH (owned by a family member of GS)

In the inflamed PD-L1⁺ patients (G1.1), spatial analysis revealed that an elevated intratumoral CD68⁺CD163⁺ M2 macrophages as well as CD11c⁺ dendritic cell infiltration (p<0.001 each) was associated with a high (CD3⁺CD4[±]CD8[±]FOXP3[±]) T-cell exclusion and a high PD-1 expression on T-cells (p<0.001 each).

In breast cancer, PD-L1 intensity on cells tumor showed a higher predictive performance for overall survival with area an under receiver operating curves (AUC) of 0.72 (p<0.0001) than percentage of PD-L1⁺ tumor cells (AUC: 0.54)

> The highly multiplexed fluorescence IHC BLEACH&STAIN framework for automated PD-L1, PD-1 and CTLA-4 assessment revealed that PD-L1 positive tumor samples can be also associated with a noninflamed TME and high spatial T-cell exclusion.

- \geq 6 major PD-L1 phenotypes were identified ranging from:
- an inflamed PD-L1⁺ tumor cell phenotype with a **spatial T-cell exclusion** (G1.1)
- to a non-inflamed PD-L1⁺ immune cell phenotype showing a **particular poor prognosis** (G2.2)
- to a non-inflamed PD-L1⁻ negative phenotype (G3.2).



Universitätsklinikum Hamburg-Eppendorf

Conclusions